

Safety Evaluation of *Ziziphus Mauritiana* Extract for its Utilization in Bread From Cholistan Desert, Pakistan

Tanveer Ahmad^{*a}, Muhammad Madnee^{*b}, Zulfiqar Ahmad^a, Muhammad Abid^b Muhammad Ammar Khan^a, Nugraha Akbar Nurrochmat^c

^aDepartment of Food Science and Technology, The Islamia University of Bahawalpur, Pakistan

^bInstitute of Forest Sciences, The Islamia University of Bahawalpur, 63100, Pakistan

^cForestry Discipline, Field of Agricultural Science, Doctoral School, Warsaw University of Life Sciences, Warsaw 02-787, Poland

***Corresponding authors:** jamtanveer70@gmail.com, muhammad.madnee@iub.edu.pk

Abstract: Considering health risk potential of various chemical additives in food, a study was conducted to evaluate the safety assessment potential of *Z. mauritiana* extract to be used in the bread manufacturing process. Mature and fresh leaves were subjected to drying at 60°C for ethanol extraction. Various dosages of extract were provided to experimental rats for safety assessment. Blood samples were evaluated based on different parameters; liver working tests, kidney function tests, lipid profile, cardiac enzymes and serum proteins. Results explicated the non-remarkable influence of concentration on different characteristics of blood. ALT concentrations express the non-momentous and variable influence of concentrate. The plant is recommended for sustainable utilization and consumption in the food and pharmaceutical industry.

Keywords: *Ziziphus mauritiana*, Safety Evaluation, Liver, Kidney functions, Food

Article History

Received: 29 January 2024; Revised: 28 February 2024; Accepted: 14 March 2024

Online: 21 April 2024; Published: Volume (3) Issue (1) May 10, 2024

<https://doi.org/10.69501/jgjmr487>

1. Introduction

Ziziphus mauritiana is a small tree or shrub that grows in drought conditions (Awasthi and More, 2009). Its plants are dispersed in sub-tropical and warm areas of the world. It is an indigenous plant of the Indo-Pak subcontinent and China (Bhargava *et al.*, 2005). It is also grown in Australia, Russia, Southern Europe, the Middle East, the USA and Asian countries like Pakistan, India, China, Thailand, Vietnam, and Korea (Reich, 1991; William, 2002). It is an extensively growing plant and is known by different local names (Upadhyay *et al.*, 2012).

In Pakistan, *Z. mauritiana* producing areas are Bahawalpur, Multan, Faisalabad, Sargodha, Jhang (Punjab province), Hyderabad, Tandojam, Nawabshah, Mirpurkhas, Gadop and Malir Area of Karachi (Sindh). The average production of fruit per tree is 80-200 kg depending upon climatic conditions (Mukhtar *et al.*, 2004). The desert of Cholistan is spread over an area of 26,000 km². Its location is Southern Punjab, Pakistan (Rao *et al.*, 1989; Arshad *et al.*, 2007). The climate of Cholistan is sub-tropical and is suitable for the growth of *Z. mauritiana*. The local people use *Z. mauritiana* to cure various diseases (Arshad *et al.*, 2006;2007). The usage of medicinal herbs for treating ailments is as old as mankind. These plants have gotten more

attention from research centers because of their importance for the safety of humanity (Najafi *et al.*, 2010). The medicinal benefits of this vegetation are because of several bioactive compounds (Karthikeyan *et al.*, 2003).

The major compounds present in *Z. mauritiana* are lipids, carbohydrates, alkaloids, saponins, triterpenic acid and glycosides. The leaves contain various compounds like triterpenic acid, volatile oils, glycosides, saponins and flavonoids. The phenolic compounds have therapeutic effects such as antithrombotic, antibacterial, anti-allergic, antiviral, antimutagenic and antineoplastic. These effects are due to antioxidant substances and free radical scavenging potentials of various polyphenolic compounds present in it (Forster *et al.*, 2003; Lee *et al.*, 2004).

The concentrates of leaves, seeds and fruit of *Z. mauritiana* show properties to slow down the process of oxidation (Ndhalala *et al.*, 2006; Bhatia and Mishra, 2009). The bark and pulp extracts show cytotoxicity against cancer cells (Ramadoss *et al.*, 2000; Vahedi, 2008). The fruits of *Z. mauritiana* are used to cure various ailments such as healing of wounds, pulmonary disorders and fever. These are used as active ingredients of Joshanda formulation, which are effective against chest problems. These also act as blood purifiers

and digestants. The dried fruits have laxative effects (Adzu *et al.*, 2003; Goyal *et al.*, 2012).

The alcoholic concentrate of *Z. mauritiana* roots inhibits diarrhea (Adzu *et al.*, 2003).. The powder of roots is applied to wounds for quick healing (Goyal *et al.*, 2012). Pectin present in *Z. mauritiana* has bile acid binding, cholesterol-lowering and antidiarrheal properties (Xiao *et al.*, 2013). Betulinic acid present in *Z. mauritiana* is a pentacyclic triterpene and lipophilic compound that has cytotoxic effects against cancer cell (Zhang *et al.*, 2009). It also inhibits enzyme aminopeptidase-N, which is involved in the regulation of angiogenesis (Kwon *et al.*, 2002).

Goyal *et al.* (2012) reported that seeds have calming and sedative effects. They have the potential to stop vomiting and nausea. They also provide relief against abdominal pain during pregnancy. According to Jarald *et al.* (2009), various extracts of *Z. mauritiana* fruits have anti-hyperglycemic potential. They also lower serum cholesterol, glucose, creatinine, triglyceride, urea, and glycosylated hemoglobin and improve HDL levels. The methanolic extract of *Z. mauritiana* has antioxidant and anti-diarrheal activities. The fruits have a hepato-protective effect (Dahiru *et al.*, 2006; Ndhlala *et al.*,

2008). The seeds contain a volatile oil that promotes hair growth (Yoon *et al.*, 2010).

Concentrate of *Z. mauritiana* inhibits the growth of liver tumor cells, malignant tumor cells and fibroblastic cells (Fu *et al.*, 2005; Kumar *et al.*, 2010). The methanolic extract shows antimicrobial effects against various bacterial and fungal strains. It also has antagonistic effects against Gram-negative and Gram-positive bacteria (Kayser and Arndt, 2000). Betulinic acid is an active compound isolated from *Z. mauritiana*. It inhibits the urease activity of *Helicobacter pylori* (Shin *et al.*, 2009). Rutin minimizes resistance in airways (Jung *et al.*, 2007). Rutin prevents osteopenia by increasing the osteoplastic activity of bones. It can also be known as an osteoblast stimulant (Molteni *et al.*, 2000). People are nowadays more conscious of the health risks of chemical additives in foods. Therefore, Considering the health risk potential of various chemical additives in food, a study was conducted to evaluate the safety assessment potential of *Z. mauritiana* extract to be used in the bread manufacturing process.

2. Materials and Methods

Raw material (foliage of *Ziziphus mauritiana*) was collected from Cholistan. The necessary chemical substances were purchased from the supplier. After the

collection of leaves, washing and cleaning were carried out to remove dust particles. The moisture from the leaves was removed in the sun and then oven (hot air oven) dried at 60°C for sixty minutes. Later, they were crushed finely and sieved to remove coarse particles. The ground sample was put in a soxhlet extraction apparatus in the presence of ethanol. After the evaporation of ethanol, the ethanolic extract was recovered and stored at 4°C till further analysis.

2.1. Safety Assessment of concentrate

Before the application of antioxidant concentrate in food products, its safety was evaluated using Sprague Dawley rats. Various treatments like $T_1=0.5$, $T_2=1.0$, $T_3=1.5$ and $T_4=2$ g/kg body weight/day of plant extract together with T_0 (control group) were administered through oral cavity to different sets of study animals for two months. The blood specimens were collected and different parameters like bilirubin, AST, ALT (liver working tests), urea, creatinine (kidney function tests), cholesterol, triglycerides, HDL, LDL (lipid profile), LDH, CPK, CKMB (cardiac enzymes), total protein and albumin (serum proteins) were determined spectrophotometrically by means of kits (Hussain *et al.*, 2012; Ahmad *et al.*, 2013).

3. Results and discussion

3.1. *Ziziphus mauritiana* concentrate

Plant concentrate was gained using the soxhlet apparatus. Ethanol was used as a solvent. After the vaporization of ethanol, 1.30 ± 0.10 % plant extract was obtained. Further assessment showed total polyphenols 25.92 ± 1.51 mg(GAE)/g, antioxidants 6.30 ± 0.32 g/ml and DPPH assay (free radicals scavenging activity) 52.75 ± 5.49 mg/ml.

3.2. Safety assessment of *Z.mauritiana* extract

The safety of concentrate was assessed, employing different treatments such as $T_1=0.5$, $T_2=1.0$, $T_3=1.5$ and $T_4=2$ g/kg.bw/day of plant concentrate to different sets of experimental animals (Sprague Dawley rats) for 2 months along with T_0 (control group). The rats were grouped in five sets with four rats in each set. Throughout the study period, no animal showed any symptoms of health complications. Later, the samples of blood were taken from experimental animals and studied concerning liver and kidney function tests, cardiac enzymes, lipid profile and serum proteins.

3.3. Liver function tests

Mean values (Table 01) explain the momentous influence of various treatments of concentrate on bilirubin, AST and ALT levels of blood in experimental rats. Data concerning bilirubin concentration shows



Fig. 01 (*Z. mauritiana*) A. Whole plant B. Fruit C. Flowers and leaves D. Seeds

non-momentous and arbitrary influence of different treatments of concentrate. The lot T₂ of rats exhibited maximum concentration (0.49 ± 0.02^a mg/dL), whereas the concentrations of 0.48 ± 0.02^a , 0.47 ± 0.02^a and 0.46 ± 0.02^a mg/dL were shown by groups T₁, T₀ and T₄ correspondingly. While the lot T₃ of study animals revealed the lowest (0.44 ± 0.02^a mg/dL) value. In the case of AST, the group T₃ of rats showed the maximum (45.63 ± 1.88^a U/L) value, whereas the values of 44.50 ± 1.34^a , 43.50 ± 1.31^a and 43.20 ± 1.93^a U/L were expressed by the sets T₀, T₄ and T₂ of study animals correspondingly. Nevertheless, the minimum concentration of 42.34 ± 0.83^a U/L was shown by group T₁.

The results concerning ALT concentrations express the non-momentous and variable influence of concentrate. The plot T₁ of rats exhibited the minimum concentration (50.14 ± 2.83^a U/L); whereas, the mean concentrations of 51.12 ± 1.90^a , 52.79 ± 2.76^a and 53.01 ± 2.22^a were gained by the groups

T₀, T₄ and T₂ accordingly. Whereas, the group T₃ of rats expressed the maximum concentration as 54.41 ± 1.95^a U/L. From the above results, it is obvious that concentrate had no detrimental influence on liver function parameters. These findings are in line with (Kumar *et al.*, 2022).

Table 01. Effect of concentrate on liver function tests in experimental rats

Treatments	Bilirubin (mg/dL)	AST (U/L)	ALT (U/L)
T ₀	0.47 ± 0.02^a	44.50 ± 1.34^a	51.12 ± 1.90^a
T ₁	0.48 ± 0.02^a	42.34 ± 0.83^a	50.14 ± 2.83^a
T ₂	0.49 ± 0.02^a	43.20 ± 1.93^a	53.01 ± 2.22^a
T ₃	0.44 ± 0.02^a	45.63 ± 1.88^a	54.41 ± 1.95^a
T ₄	0.46 ± 0.02^a	43.50 ± 1.31^a	52.79 ± 2.76^a

*n=4, $\pm$ S.E., in columns means with the same letter are non-significant

3.4. Kidney function tests

The data (Table 02) exhibit that various dosages of concentrate exerted no remarkable impact on different parameters of the kidney. Means about urea concentration express that the category T₀ of study animals revealed the lowest concentration (29.32 ± 1.25^a mg/dL). Although the mean concentrations of

29.61±1.65^a, 30.46±1.90^a and 31.61±1.64^a mg/dL were exhibited by sets, T₁ and T₂ accordingly. Nevertheless, the highest concentration of 32.06±0.77^a mg/dL was expressed by the set T₄.

Table 02. Effect of concentrate on kidney function tests in experimental rats

Treatments	Urea (mg/dL)	Creatinine (mg/dL)
T ₀	29.32±1.25 ^a	0.62±0.01 ^a
T ₁	30.46±1.90 ^a	0.63±0.01 ^a
T ₂	31.61±1.64 ^a	0.61±0.02 ^a
T ₃	29.61±1.65 ^a	0.64±0.02 ^a
T ₄	32.06±0.77 ^a	0.65±0.03 ^a

*n=4, ± S.E., in columns means with the same letter are non-significant

The results regarding creatinine explicitly that various levels of plant extract had a minor and haphazard impact on its concentration. The rats of group T₄ exhibited maximum (0.65±0.03^a mg/dL) value, while the values of 0.64±0.02^a, 0.63±0.01^a and 0.62±0.01^a mg/dL values were shown by the sets T₃, T₁ and T₀ respectively. However, the class of experimental animals having the treatment T₂ expressed minimum concentration (0.61±0.02^a mg/dL). The findings obtained from the above parameters clear that different levels of concentrate did

not show dangerous effects on urea and creatinine concentrations of blood. These results are similar to the earlier study by Sharma et al. (2020).

3.5. Cardiac enzymes

The findings (Table 03) expound the minute influence of several dosages of concentrate on LDH, CPK, and CKMB levels of blood. The outcomes for LDH show that the set T₀ of rats expressed minimum mean concentration as 161.75±4.07^aU/L, whereas mean concentrations of 162.63±4.95^a, 163.25±4.56^a and 163.75±2.64^aU/L were revealed by the lots T₂, T₁ and T₄ respectively. Nonetheless, the group of rats that received the treatment T₃ depicted a value of 164.25±3.82^a U/L. The results concerning CPK explicated that the highest value of 35.33±1.79^a was obtained by set T₄ of rats; while, the values of 34.11±1.12^a, 33.24±1.23^a and 32.24±1.13^a were expressed by the sets T₂, T₀ and T₃ accordingly. Although the rats of class T₁ exhibited the lowest (31.88±1.15^aU/L) value.

In the case of CKMB, the findings reveal that the set T₀ exhibited the least value of 82.10±4.00^a followed by the mean values of 83.20±3.03^a, 84.73±2.84^a and 85.86±1.76^a for the groups T₂, T₁ and T₄ accordingly. Nevertheless, the set T₃ revealed the highest concentration (86.06±4.33^a). The above outcomes show that various treatments of

Table 03. Effect of concentrate on cardiac enzymes in experimental rats

Treatments	LDH	CPK	CKMB
T ₀	161.75±4.07 ^a	33.24±1.23 ^a	82.10±4.00 ^a
T ₁	163.25±4.56 ^a	31.88±1.15 ^a	84.73±2.84 ^a
T ₂	162.63±4.95 ^a	34.11±1.12 ^a	83.20±3.03 ^a
T ₃	164.25±3.82 ^a	32.24±1.13 ^a	86.06±4.33 ^a
T ₄	163.75±2.64 ^a	35.33±1.79 ^a	85.86±1.76 ^a

*n=4, ± S.E., in columns means with the same letter are non-significant

Table 04. Effect of *Z. mauritiana* concentrate on lipid profile in experimental rats

Treatments	Cholesterol (mg/dL)	Triglycerides (mg/dL)	LDL (mg/dL)	HDL (mg/dL)
T ₀	95.38±3.13 ^a	85.01±1.81 ^a	36.00±1.22 ^a	40.35±2.47 ^a
T ₁	97.23±3.69 ^a	81.44±4.38 ^a	33.91±0.87 ^a	39.80±1.21 ^a
T ₂	94.88±2.46 ^a	83.03±2.26 ^a	34.27±1.73 ^a	38.41±2.09 ^a
T ₃	92.14±1.87 ^a	79.41±4.06 ^a	33.00±1.91 ^a	37.48±0.80 ^a
T ₄	90.64±3.76 ^a	77.38±2.98 ^a	35.11±1.20 ^a	38.70±0.87 ^a

*n=4, ± S.E., in columns means with the same letter are non-significant

concentrate had noxious effects on different cardiac enzymes of blood. These results coincide with an earlier study by Olaleye et al. (2019).

3.6. Lipid profile

Mean values (Table 04) explicate the minute influence of various levels of concentrate on cholesterol, triglycerides, LDL and HDL concentrations of rats. Results concerning cholesterol levels express that the group T₄ of rats exhibited the minimum value (90.64±3.76^a mg/dL), though the concentrations of 92.14±1.87^a, 94.88±2.46^a and 95.38±3.13^a mg/dL were explicated by the sets T₃, T₂ and T₀ correspondingly. However, the maximum concentration (97.23±3.69^a mg/dL) was expressed by set T₁

of rats. The results regarding triglycerides exhibited the maximum concentration (85.01±1.81^a mg/dL) for the control group (T₀), whereas the groups T₁, T₂ and T₃ of rats exhibited the mean concentrations as 81.44±4.38^a, 83.03±2.26^a and 79.41±4.06^amg/dL accordingly. However, the lowest mean concentration (77.38±2.98^a mg/dL) was obtained by a set T₄ of rats.

The outcomes concerning LDL concentration expound that the set T₀ of rats expressed maximum concentration (36.00±1.22^amg/dL), while the values of 35.11±1.20^a and 34.27±1.73^a mg/dL were showed by the sets T₄ and T₂ accordingly. Whereas, the lot T₃ of study animals exhibited the lowest mean concentrations

33.00±1.91^a mg/dL. The outcomes regarding the effect of different levels of plant concentrate on LDL levels reveal that these did not cause any harmful effects in experimental animals.

Table 05. Effect of concentrate on serum proteins in experimental rats

Treatments	Total Protein (g/dL)	Albumin (g/dL)
T ₀	5.93±0.26 ^a	2.40±0.14 ^a
T ₁	6.16±0.42 ^a	2.45±0.10 ^a
T ₂	5.95±0.09 ^a	2.56±0.14 ^a
T ₃	6.14±0.06 ^a	2.55±0.10 ^a
T ₄	5.99±0.23 ^a	2.34±0.54 ^a

Means about HDL concentrations show that the highest value of 40.35±2.47^a mg/dL was showed by set T₀ of rats, though the sets T₁, T₂ and T₃ revealed mean values as 39.80±1.21^a, 38.41±2.09^a and 37.48±0.80^a mg/dL respectively. However, the value of 38.70±0.87^a mg/dL was gained by set T₄ of rats. The above data explicates that various levels of concentrate had no detrimental influence on the lipid profile of experimental animals. These results are similar to earlier work done by Singh et al. (2018).

3.7 Serum proteins

The results (Table 05) expound that various treatments of concentrate had non-significant effects on serum proteins. The outcomes regarding total protein explicitly that the set

T₀ expressed the minimum (5.93±0.26^a g/dL) value. Whereas, the values of 5.95±0.09^a, 5.99±0.23^a and 6.14±0.06^a g/dL were shown by the sets of rats that received treatments T₂, T₄ and T₃ respectively. However, the maximum mean concentration of 6.16±0.42^a g/dL was showed by the set T₁ of rats.

The results concerning albumin (Table 05) express that the set T₀ of rats exhibited the mean concentration as 2.40±0.14^a g/dL, while the treatments T₁, T₂ and T₃ revealed concentrations as 2.45±0.10^a, 2.56±0.14^a and 2.55±0.10^a g/dL accordingly. Nevertheless, the class T₄ of experimental animals showed mean concentrations of 2.34±0.54^a g/dL. The above findings show, that concentrate did not exhibit a detrimental impact on serum proteins of blood. According to the given results, it is clear that various levels of plant concentrate did not exert a harmful impact on different parameters (liver and kidney function tests, cardiac enzymes, lipid profile and serum proteins) of the blood of Sprague Dawley rats. Therefore, it may be utilized in various foods. These results are earlier stated by Kumar et al. (2017).

4. Conclusion

Z. mauritiana concentrate has various medicinal benefits due to the presence of flavonoids, triterpinic acids, alkaloids, saponins and glycosides. The extract showed no toxic effect on bilirubin concentrations in

blood. Regarding the outcomes of AST and ALT levels, the values were also statistically non-significant. The outcomes concerning the kidney function test exhibited a non-remarkable impact of the ethanolic extract on urea and creatinine concentrations of blood. As far as the results of cardiac enzymes, different treatments of plant extract did not show harmful effects. Different parameters of lipid profile and serum proteins also expressed the non-momentous impact of extract. It is safe for humans to use this plant extract in various products. Therefore, our study recommends its utilization in various products of food items and pharmaceutical drugs. Huge potential exists for plant extract utilization in food items discouraging chemical additives that pose health risks. Further studies are needed for the exploration of the flora of Cholistan for various useful additives replacing chemical additives in food.

Acknowledgment

The authors are thankful to Professor Dr. Tanveer Hussain for providing the necessary facilities in the phytochemical laboratory for plant extract analysis and identification.

References

Ahmad, Z., M.S. Butt, R. Hussain, A. Ahmed and M. Riaz. 2013. Effect of oral application of xylanase on several blood

- parameters in broilers. *Pak Vet J.* 33(3): 388-390.
- Adzu, B., S. Amos, M.B. Amizan and K. Gamaniel. 2003. Evaluation of the antidiarrhoeal effects of *Zizyphus spina-christi* stem bark in rats. *Actatropica.* 87: (2), 245-250.
- Arshad, M., M. Ashraf and N. Arif. 2006. Morphological variability of *Prosopis cineraria* Druce, from the Cholistan desert, Pakistan. *Genet. Resour. Crop Evol.* 53:1589-1596.
- Arshad, M., M.Y. Ashraf, M. Ahamad and F. Zaman. 2007. Morpho-genetic variability potential of *Cenchrus ciliaris* L. from Cholistan desert, Pakistan. *Pak. J. Bot.* 39: 1481-1488.
- Awasthi, O. P. and T.A. More. 2009. Genetic diversity and status of *Ziziphus* in India. *Acta Hort.* 840: 33-40.
- Bhargava, R., A.K. Shukla, N. Chauhan, B.B. Vashishtha and D.G. Dhardar. 2005. Impact of hybridity on flavonoid spectrum of ber (*Z. mauritiana*). *Environ. Exp. Bot.* 53: 135-138.
- Bhatia, A. and T. Mishra. 2009. Free radical scavenging and antioxidant potential of *Z. mauritiana* seed extract. *Journal of Complementary and Integrative Medicine.* 8: 42-46.
- Dahiru, D., J.M. Sini and A.L. John. 2006. Antidiarrhoeal activity of *Z. mauritiana* root extract in rodents. *Afr J Biotechnol.* 5(10):941-945.
- Forster, M.P., M.V. Mendez, C.R. Delgado, R.E.M. Rodriguez and C.D. Romero. 2003. *Eur Food Res Technol.* 1007: 451-458.
- Fu, L., S. Zhang, N. Li, J. Wang, M. Zhao, J. Sakai, T. Hasegawa, T. Mitsui, T. Kataoka, S. Oka, M. Kiuchi, K. Hirose and M. Ando. 2005. Three New Triterpenes from *Nerium oleander* and Biological Activity of the Isolated Compounds. *Journal of Natural Products.* 68 (2): 198-206.
- Goyal, M., B.P. Nagori and D. Sasmal. 2012. Review on ethnomedicinal uses,

- pharmacological activity and phytochemical constituents of *Z. mauritiana* (*Z. jujuba* Lam., non Mill). *Spatula DD.* 2(2), 107-116.
- Hussain, R., F. Mahmood, A. Khan, M.T. Javed, S. Rehan and T. Mehdi. 2012. Cellular and biochemical effects induced by atrazine on blood of male Japanese quail (*Coturnix Japonica*). *Pesticide Biochemistry and Physiology.* 103:38-42
- Molteni, M.N.H., V.Crespy,V. Coxam, M.J.Davico, C.Rémésy, J.P.Barlet. 2000. Rutin inhibits ovariectomy-induced osteopenia in rats. *J Bone Miner Res.*15(11): 2251-2258.
- Jarald, E. E., Joshi, S. B., & Jain, D. C. (2009). Antidiabetic activity of extracts and fraction of *Zizyphus mauritiana*. *Pharmaceutical biology*, 47(4), 328-334.
- Jung, C.H., J.Y. Lee, C.H. Cho and C.J. Kim.2007. Anti-asthmatic action of quercetin and rutin in conscious guinea-pigs challenged with aerosolized ovalbumin. *Arch Pharm Res.* 30(12), 1599-1607.
- Karthikeyan, K., B.R. Bai, K. Gauthaman, K.S. Sathish and S.N. Devaraj.2003. cardioprotective effect of the alcoholic extract of *Terminaliaarjuna* bark in an in vivo model of myocardial ischemic reperfusion injury. *Life Sciences.* 73: 2727-2739.
- Kayser, O. and S.K. Arndt. 2000. Antimicrobial activity of some *Zizyphus* species used in traditional medicine. *PharmaPharmacoloLett.* 10:38-40.
- Kumar, D., S.Mallick, J.R.Vedasiromoni and B.C. Pal. 2010.Anti-Leukemicactivity of *Dilleniaindica* L. Fruit Extract and Quantification of Betulinic Acid by HPLC.*Phytomedicine.* 17(6): 431-435.
- Kwon, H.J., J.S. Shim, J.H. Kim, H.Y. Cho, Y.N. Yum,S.H. Kim. 2002. Betulinic acid inhibits growth factor-induced in vitro angiogenesis via the modulation of mitochondrial function in endothelial cells. *Jpn J Cancer Res.* 93: 417-25.
- Lee M.F., Y.H. Chen, J.L.Lan, C.Y. Tseng and C.H. Wu2004. Allergenic components of Indian jujube (*Zizyphus mauritiana*) show IgE cross-reactivity with latex allergen. *Int Arch AllerImmunol.* 133 (3):211-16.
- Mukhtar, H.M., S.H. Ansari,M. Ali and T.Naved. 2004. New compounds from *Zyzyphus vulgaris*. *Pharmaceutical Biology.* 42: 508-511.
- Najafi, S., N. Sanadgol, B.Sadeghinejad, B.Ashofteh. 2010. Phytochemical sreening and antibacterial activity of *Citrullus colocynthis*Schrad against *Staphylococcus aureus*. *JOMPR.*4(9) 2321-2325.
- Ndhala. A.R.,C.H. Mupure, K.Chitindingue. M.A.N.Benhura and M.Muchuweti.2006. Antioxidant potentials and degree of polymerization of six wild fruits. *Life Science Research Assays.* 1: 87-92.
- Ndhala, A.R., K.Chitindingu, C.Mupure,T. Murenje, F. Ndhalaand M.A.Benhura. 2008. Antioxidant properties of methanolic extracts from *Diospyromespiliformis* (jackal berry), *FlacourtiaindicaZ. mauritiana* (Batoka plum), *Uapacakirkiana* (wild loquat) and *Z. mauritiana* (yellow berry) fruits. *Int J Food Sci Tech.* 43:284-8.
- Oluwasina, O. O., Olaleye, F. K., Olusegun, S. J., Oluwasina, O. O., & Mohallem, N. D. (2019). Influence of oxidized starch on physicommechanical, thermal properties, and atomic force micrographs of cassava starch bioplastic film. *International journal of biological macromolecules*, 135, 282-293.
- Ramadoss, S., M. Jaggiand M.J.A.Siddiqui. 2000. Use of betulinic acid and its derivatives for inhibiting cancer growth and a method of monitoring this. *Assignee: Dabur Research Foundation.* US patent No.6048847.

- Rao, A.R., M. Arshad and M. Shafiq. 1989. Perennial Grass Germplasm of Cholistan Desert and its Phytosociology. *Cholistan Institute of Desert Studies, Islamia University, Bahawalpur, Pakistan*; 160.
- Reich, L. 1991. Uncommon fruits worthy of attention. *A Gardener's Guide*. Addison Wesley, Reading, M. A.
- Shin, S., C.E. Park, N.I. Baek, I.C. Chung and C.H. Park. 2009. Betulinic and Oleanolic Acids Isolated from *Forsythia suspense* Vahl Inhibit Urease Activity of *Helicobacter pylori*. *Biotechnology and Bioprocess Engineering*. 14(2): 140-145.
- Sharma V, Sharma R, Gautam DS, Kuca K, Nepovimova E, Martins N. Role of Vacha (*Acorus calamus* Linn.) in Neurological and Metabolic Disorders: Evidence from Ethnopharmacology, Phytochemistry, Pharmacology and Clinical Study. *J Clin Med*. 2020 Apr 19;9(4):117
- Upadhyay, S., P. Upadhyay, A.K. Ghosh and V. Singh. 2012. *Z. mauritiana*: a review on pharmacological potential of this underutilized plant. *international journal of current Research and Review*. 04(03).
- Vahedi, F., Fathi, M. Najafi and K. Bozari. 2008. Evaluation of inhibitory effect and apoptosis induction of *Zyzyphus Jujube* on tumor cell lines, an in vitro preliminary study," *Cytotechnology*; 56 (2) 105–111.
- William, H. (2002). Notes on economic plants. *Economic Botany*. 56:198-206.
- Xiao, H.T., S.W. Tsang, H.Y. Qin, F.F. Choi, Z.J. Yang, Q.B. Han and Z.X. Bian. 2013. A bioactivity-guided study on the anti-diarrheal activity of *Polygonum chinense* Linn. *Journal of ethnopharmacology*; 149(2): 499-505.
- Yoon, J.I., M. Sharif, Al-Reza and S.C. Kang. 2010. Hair growth promoting effect of *Zizyphus jujuba* essential oil. *Food and Chemical Toxicology*. 48: 1350–1354.
- Zhang, Y.X., C.Z. Kong, H.Q. Wang, L.H. Wang, C.L. Xu and Y.H. Sun. 2009. Phosphorylation of Bcl-2 and activation of caspase-3 via the c-Jun N-terminal kinase pathway in ursolic acid-induced cells apoptosis. *Biochimie*; 91:1173–9.